

INHERITANCE OF RESISTANCE TO A HIGHLY VIRULENT RACE F OF *Orobanche cumana* Wallr. IN A SUNFLOWER LINE DERIVED FROM INTERSPECIFIC AMPHIPLOIDS

B. Pérez-Vich¹, B. Akhtouch¹, J. Muñoz-Ruz¹,
J.M. Fernández-Martínez¹ and C.C. Jan^{2*}

¹Instituto de Agricultura Sostenible (CSIC), Apartado 4084,
E-14080 Córdoba, Spain

²Northern Crop Science Laboratory, USDA-ARS, Box 5677,
Fargo, ND 58105, USA

Received: November 08, 2001

Accepted: June 03, 2002

SUMMARY

Broomrape (*Orobanche cumana* Wallr.) populations belonging to the new race F in Spain have overcome all known resistance genes, Or_1 to Or_5 , in cultivated sunflower (*Helianthus annuus* L.) and are spreading rapidly. Resistance to race F of this parasitic weed has been found in wild perennial species of *Helianthus* and has been introgressed into cultivated sunflower. The objective of this study was to characterize the inheritance of resistance genes in cultivated sunflower derived from wild perennial species *H. divaricatus* and *H. grosseserratus*, respectively. Crosses between resistant cultivated lines and the susceptible line P21 were made, and the F_1 's were resistant when evaluated for broomrape resistance using a highly virulent population of race F, indicating dominance of resistance genes. Comparison of resistance of the segregating populations, F_2 and BC_1F_1 , to both parents confirmed the dominance observed in the F_1 and indicated that resistance is under the control of a single dominant gene. This dominance of resistance genes will greatly simplify the breeding for resistance.

Key words: sunflower, *Helianthus annuus* L., inheritance, broomrape, *Orobanche cumana* Wallr., race F

INTRODUCTION

Sunflower broomrape (*Orobanche cumana* Wallr.) is a parasitic angiosperm, totally devoid of chlorophyll, that infects the roots of sunflower (*Helianthus annuus* L.) plants, drawing water and nutrients from them. This parasitic plant is regarded

* Corresponding author: Phone: +1 701 239 1319,
Fax: +1 701 239 1346, e-mail: Janc@fargo.ars.usda.gov

as one of the most important constraints on sunflower production in areas of eastern and southern Europe, the Middle East, Russia, Ukraine and China (Parker, 1994). Broomrape attacks are frequently severe and yield losses have reached up to 50% (Domínguez, 1996a). Though several methods of control have been proposed, herbicide control is only partially effective (García-Torres *et al.*, 1988) and soil treatments by fumigation or solarization are effective but not economically feasible (Foy *et al.*, 1989; Jacobsohn *et al.*, 1987). The most economical and effective means of controlling sunflower broomrape is the use of resistant cultivars.

Genetic resistance to broomrape has been introduced into sunflower cultivars from early sunflower breeding programs in the former USSR (Pustovoit, 1966). However, the widespread use of resistant cultivars has led to the appearance of new races of the parasite that overcome the resistance genes (Škorić, 1988), and a continuous need for new resistance sources. Cultivated sunflower has a narrow genetic background and is deficient in genes for resistance to *O. cumana* and other pathogens and pests. The main sources of resistance to *O. cumana* in sunflower have been identified in wild *Helianthus* species (Pustovoit, 1966; Korell *et al.*, 1996; Ruso *et al.*, 1996; Fernández-Martínez *et al.*, 2000).

The host-parasite system of sunflower-*O. cumana* appears to follow the gene-for-gene model. Vrânceanu *et al.* (1980) established a set of five sunflower differentials carrying the five dominant resistant genes *Or*₁ to *Or*₅, each resistant to a new race and also to the previous race. These differential lines along with a universal susceptible host permitted the identification of five physiological races designated A to E. Genetic mechanisms controlling resistance to broomrape proposed more recently by other authors agreed with these results, and monogenic and dominant inheritance of resistance to sunflower broomrape was described (Pogorletsky and Geshele, 1975; Ish-Shalom-Gordon, 1993; Sukno *et al.*, 1999). Domínguez (1996b) identified one line carrying two of these dominant resistance genes.

Orobanche cumana is known to have been present in Spain since 1958. Melero-Vara *et al.* (1997) identified races that overcame *Or*₁, *Or*₃, and *Or*₄ genes, but not *Or*₂ and *Or*₅. However, recent studies have shown an evolution of sunflower broomrape races in Spain, with a new race, designated as F, overcoming all the resistance genes identified so far (Alonso *et al.*, 1996; Domínguez, 1999). Resistance to this new race has been found in cultivated and in wild sunflower (Sukno *et al.*, 1999; Domínguez, 1999; Fernández-Martínez *et al.*, 2000). Rodríguez-Ojeda *et al.* (2001) and Akhtouch *et al.* (2001) identified two independent recessive genes controlling resistance to race F in lines derived from cultivated germplasm.

The high number of immune entries resistant to broomrape race F observed in populations of wild perennial sunflower (Fernández-Martínez *et al.*, 2000) prompted the initiation of a program to transfer the resistance genes into cultivated sunflower. As a first result, the four germplasm populations, BR1 to BR4, resistant to race F and derived from the wild perennial sunflower species *H. maximiliani*, *H. grosseserratus*, and *H. divaricatus*, were released (Jan *et al.*, 2001). In order to

use efficiently the available sources of broomrape resistance, it is necessary to determine the inheritance of resistance. The objective of this study was to determine the inheritance of resistance to broomrape race F in material derived from BR4.

MATERIALS AND METHODS

Plant materials

The plant materials used for the genetic study were:

1. an inbred line developed from BR4, which is a population resistant to race F of broomrape and derived from the sunflower wild perennial *H. divaricatus* and *H. grosseserratus* (Jan *et al.*, 2001);
2. P21, a cultivar selected from "Peredovik" carrying a recessive gene for male sterility and highly susceptible to broomrape races E and F; and
3. NR5, a cultivated line of sunflower carrying the resistance gene *Or₅*.

The pedigree of BR4 is as follows: *H. divaricatus*-830/P21,D//*H. grosseserratus*-001/P21,D/3/P21/4/HA89. Chromosomally doubled (D) F₁ heads of *H. divaricatus*-830 x P21 and *H. grosseserratus* x P21 were intercrossed and the resulting mixed-amphiploid maintained by sib-pollination. This mix-amphiploid was first backcrossed with P21, and the resulting triploid BC₁F₁'s backcrossed with HA89 to produce BC₂F₁ (Jan *et al.*, 2001). Evaluations for broomrape race F resistance of BC₂F₁, BC₂F₂, and BC₂F₃ were then performed (Jan *et al.*, 2000). The BR4 material was a BC₂F₄ bulk from 10 resistant BC₂F₃ families grown from selected immune BC₂F₂ plants. For the genetic study, the BC₂F₄ seeds were self-pollinated two times and checked again for resistance to race F of broomrape. All plants were breeding true for resistance. The BC₂F₆ material was designated as J1.

Genetic study

Crosses between resistant plants of J1, artificially inoculated with race F of broomrape, and male-sterile plants of the susceptible line P21 were carried out in the greenhouse in the winter of 2000. Plants of the parental lines J1 and P21, the F₁, and the check line NR5, which carries *Or₅*, were artificially inoculated with race F of broomrape and grown in spring-2001 at the Institute for Sustainable Agriculture at Córdoba (Southern Spain). These plants were kept outdoors until maturity. F₂ seeds were produced by self-pollinating F₁ plants, and BC₁F₁ seeds by backcrossing to both parental lines. F₂ and BC₁F₁ plants, together with plants of J1, P21, NR5 and F₁ plants, were grown in growth chambers in summer-2001 and tested for broomrape resistance (race F) in artificially infested soil.

For the spring-2001 study, artificial inoculations were performed by planting 2-d-old sunflower seedlings in small plastic pots (7 x 7 x 8 cm) containing a mixture of sand and peat (1:1, v:v) uniformly infested with 50 mg of broomrape (race F)

seed. The plants were incubated in a growth chamber for 15-20 days at 28°C/22°C (day/night) and 14 h photoperiod. Then the plants were transplanted into larger pots containing 3 l of fertilised and uninfected sand/silt/peat (2:1:2, v:v:v) soil mixture.

For the summer-2001 study, artificial inoculations were performed by planting 2-days-old sunflower seedlings in small porex pots (4 x 4 x 7 cm) with peat uniformly infested with 35 mg of broomrape (race F) seed. The plants were kept in a growth chamber till evaluation (45 days). The incubation was conducted at 28°C/22°C (day/night) and 14 h photoperiod.

Evaluation for broomrape resistance was made on mature plants in the spring-2001, and on the 45-day-old plants grown in the growth chamber in the summer-2001. Plants were uprooted and their root system carefully washed to observe any established broomrape nodules or stalks. These were indicative of susceptibility to broomrape, whereas those lacking these structures were considered resistant. Number of tested plants is indicated in Table 1. Resistant to susceptible plant ratios were used to determine genetic relationships. Chi-square tests were used to test for goodness of fit to expected genetic ratios. A single dominant gene model (3:1 for the F₂ and 1:1 for the BC₁F₁ plants) was applied to evaluate the segregation ratio for inheritance of *O. cumana* (race F) resistance.

Table 1: Resistant (R) and susceptible (S) plants of the parental lines P21 and J1, their F₁, and the F₂ and BC₁F₁ generations, segregating for resistance to race F, from their cross. NR5, which carries the *Or₅* gene, served as a susceptible check to race F of broomrape

Experiment	Generation or line	Number of plants		Ratio tested	χ^2	P
		R	S			
Spring-2001	J1	15	0			
	P21	0	15			
	P21/J1, F ₁	15	0			
	NR5	0	9			
Summer-2001	J1	9	0			
	P21	0	3			
	P21/J1, F ₁	10	0			
	P21/J1, F ₂	77	36	3:1 (R:S)	2.83	0.10
	P21/J1//P21, BC ₁ F ₁	30	35	1:1 (R:S)	0.38	0.54
	P21/J1//J1, BC ₁ F ₁	32	0			
	NR5	0	10			

RESULTS AND DISCUSSION

The sunflower parental line J1 was resistant to broomrape race F both in spring-2001 and summer-2001 studies, while all plants of the parental line P21 and NR5, which carries the *Or₅* gene, were susceptible. This verifies the reliability of the inoculation technique, and confirmed that resistant lines were uniform and

consistent for both trials. All the fifteen F_1 plants from the J1 cross to P21 tested in spring-2001, and all ten F_1 plants from the same cross tested in summer-2001 were resistant (Table 1). These results indicated a dominant gene action in the resistance to race F of broomrape in this material, which was suggested previously by Jan *et al.* (2000) in their backcrossing program to introgress resistance to race F of broomrape from wild perennial species of *Helianthus* into cultivated sunflower.

The segregation pattern of resistance to broomrape race F observed in the F_2 generation from the J1 cross to P21 fit a 3:1 (resistant:susceptible) ratio (Table 1). The data of the BC_1F_1 to the susceptible parental line P21 generation also satisfactorily fit a 1:1 (resistant:susceptible) ratio. No susceptible plants were detected in the backcrosses to the resistant parent J1 (Table 1). These data indicated that resistance to *O. cumana* race F in the J1 line is controlled by a single dominant gene. This gene was tentatively designated as Or_6 .

These results are similar to those obtained in previous genetic studies with races A to E of broomrape, which evidenced monogenic dominant inheritance for resistance to broomrape (Pogorletskii and Geshele, 1975; Ish-Shalom-Gordon, 1993; Sukno *et al.*, 1999). However, in all the inheritance studies carried out to date with race F, broomrape resistance to this race has been shown to be recessive and controlled by alleles at two loci (Rodríguez-Ojeda *et al.*, 2001; Akhtouch *et al.*, 2001). The resistant germplasm tested in those studies was derived from cultivated sunflower. The results presented in the present work imply an important change in the pattern of resistance to race F recently observed. The dominance reaction of resistance genes will greatly simplify the breeding procedure for resistance to race F, as the incorporation of genes of resistance for the development of commercial resistant hybrids need to be introduced in only one parental line. On the other hand, the monogenic inheritance allows an easy and a successful transference of the resistance into breeding lines. The different results obtained with resistance to broomrape race F in material derived from cultivated or from wild sunflower indicates that different resistance mechanisms are involved in lines derived from these different sources, and suggests the importance of the wild sunflower species as a source of unique resistant genes.

REFERENCES

- Akhtouch, B., Muñoz-Ruz, J., Melero-Vara, J., Fernández-Martínez, J. and Domínguez, J., 2001. Inheritance of resistance to race F of sunflower broomrape (*Orobanche cumana* Wallr.). Plant Breed. (submitted).
- Alonso, L.C., Fernández-Escobar, J., López, G., Rodríguez-Ojeda, M. and Sallago, F., 1996. New highly virulent sunflower broomrape (*Orobanche cernua* Loebl.) pathotype in Spain. In: M. Moreno (ed.), Advances in Parasitic Weed Research. Proc. of the 6th Int. Symposium in Parasitic Weeds, 639-644, Córdoba, Spain.
- Domínguez, J., 1996a. Estimating effects on yield and other agronomic parameters in sunflower hybrids infested with the new races of sunflower broomrape. Proc. Symposium I: Disease Tolerance in Sunflower, 118-123, International Sunflower Association, Beijing.

- Domínguez, J., 1996b. R-41, a sunflower restorer inbred line, carrying two genes for resistance against a highly virulent Spanish population of *Orobanche cernua*. *Plant Breed.*, 115: 203-204.
- Domínguez, J., 1999. Inheritance of the resistance to *Orobanche cumana* Wallr. In: Cubero J.I. (ed.), *Resistance to broomrape: The state of the art. Congresos y jornadas 51/99*, Junta de Andalucía. Consejería de Agricultura y Pesca, 139-141.
- Fernández-Martínez, J.M., Melero-Vara, J.M., Muñoz-Ruz, J., Ruso, J. and Domínguez, J., 2000. Selection of wild and cultivated sunflower for resistance to a new broomrape race that overcomes resistance of the *Or₅* gene. *Crop Sci.*, 40: 550-555.
- Foy, C.L., Jain, R. and Jacobsohn, R., 1989. Recent approaches for chemical control of broomrape *Orobanche* ssp. *Rev. Weed Sci.*, 4: 123-152.
- García-Torres, L., Castejón-Muñoz, M. and Romero-Muñoz, F., 1988. Herbicidal selection for broomrape (*Orobanche cernua*) control in sunflower. *Helia*, 11: 65-68.
- Ish-Shalom-Gornon, N., Jacobsohn, R. and Cohen, Y., 1993. Inheritance of resistance to *Orobanche cumana* in sunflower. *Phytopathology*, 83: 1250-1252.
- Jacobsohn, R., Greenberger, A., Katan, J., Levi, M. and Alon, H. 1980. Control of Egyptian broomrape (*Orobanche egyptiaca*) and other weeds by means of solar heating of the soil by polyethylene mulching. *Weed Sci.*, 28: 312-316.
- Jan, C.C., Ruso, J., Muñoz-Ruz, J. and Fernández-Martínez, J.M., 2000. Resistance of sunflower (*Helianthus*) perennial species, interspecific amphiploids, and backcross progeny to broomrape (*O. cumana* Wallr.) races. *Proc. 15th international Sunflower Conference*, 12-15 June, 2000, Toulouse, France, pp. J-14-J-19.
- Jan, C.C., Fernández-Martínez, J.M., Ruso, J. and Muñoz-Ruz, J., 2001. Registration of four sunflower germplasm populations resistant to broomrape race F. *Crop Sci.*, (submitted).
- Korrell, M., Brahm, L., Friedt, W. and Horn, R., 1996. Interspecific and intergeneric hybridization in sunflower breeding. II Specific uses of wild germplasm. *Plant Breed. Abstr.*, 66: 1081-1091.
- Melero-Vara, J.M., 1997. El jopo del girasol: Evolución racial y desarrollo de resistencia genética. *Proc. 4th Nat. Seed Symposium*, 5-7 Nov, 1997, Sevilla, Spain, pp. 373-382.
- Parker, C., 1994. The present state of *Orobanche* problem. In: A.H. Pieterse, J.A.C. Verkleijand, and S.J. Ter Borg (eds.), *Biology and management of Orobanche*. *Proc. 3rd Int. Workshop on Orobanche and related Striga research*, Royal Tropical Institute, Amsterdam, pp. 17-26.
- Pogorletsky, P.K. and Geshele, E.E., 1976. Sunflower immunity to broomrape, and rust. *Proc. 7th Int. Sunflower Conf.*, 27 June-3 July 1976, Krasnodar, Russia, pp. 238-243.
- Pustovoit, V.S., 1966. Selection, seed culture and some agrotechnical problems of sunflower. Translated from Russian in 1976 by Indian National Scientific Documentation Centre, Delhi, India.
- Rodríguez-Ojeda, M.I., Fernández-Escobar, J. and Alonso, L.C., 2001. Sunflower inbred line (KI-374) carrying two recessive genes for resistance against a highly virulent Spanish population of *Orobanche cernua* Loelf. / *O. cumana* Wallr. race F. *Proc. 7th Int. Parasitic Weed Symposium*, 5-8 June 2001. Nantes, France, pp. 208-211.
- Ruso, J., Sukno, S., Domínguez-Jiménez, J., Melero-Vara, J.M. and Fernández-Martínez, J.M., 1996. Screening of wild *Helianthus* species and derived lines for resistance to several populations of *Orobanche cernua*. *Plant Dis.*, 80: 1165-1169.
- Saavedra del Río, M., Fernández Martínez, J.M. and Melero-Vara, J.M., 1994. Virulence of populations of *Orobanche cernua* Loelf. attacking sunflower in Spain. In: A.H. Pieterse, J.A.C. Verkleijand, and S.J. Ter Borg (eds.), *Biology and management of Orobanche*. *Proc. 3rd Int. Workshop on Orobanche and related Striga research*, Royal Tropical Institute, Amsterdam, pp. 139-141.
- Škorić, D., 1988. Sunflower breeding. *Uljarstvo*, 25: 1-90.
- Sukno, S., Melero-Vara, J.M. and Fernández-Martínez, J.M., 1999. Inheritance of resistance to *Orobanche cernua* Loelf. in six sunflower lines. *Crop Sci.*, 39: 674-678.
- Vrânceanu, A.V., 1977. El girasol. Ed. Mundi Prensa, Madrid.
- Vrânceanu, A.V., Tudor, V.A., Stoenescu, F.M. and Pirvu, N., 1980. Virulence groups of *Orobanche cumana* Wallr., differential hosts and resistance source genes in sunflower. *Proc. of the 9th Int. Sunflower Conf.*, 8-9 June, 1980. Torremolinos, Spain, 74-82.

HERENCIA DE LA RESISTENCIA A LA RAZA ALTAMENTE AGRESIVA F DE *Orobanche cumana* Wallr. EN LA LINEA DEL GIRASOL DERIVADA DE INTERSPECIES AMFIPLOIDE

RESUMEN

Las poblaciones de orobanque (*Orobanche cumana* Wallr.) que pertenecen a la raza nueva F en España no reaccionan a todos los genes de resistencia conocidos, de Or_1 a Or_5 , siendo presentes en el girasol cultivado (*Helianthus annuus* L.) y se extienden rápidamente. La resistencia a la raza F de esta mala hierba fue encontrada en las especies silvestres del girasol y transferida en el girasol cultivado. El objetivo de este trabajo era de describir la herencia de los genes de resistencia en el girasol introducidos de las especies de girasol de varios años *H. divaricatus* y *H. grosseserratus*. Las líneas resistentes del girasol cultivado fueron cruzadas con la línea sensible P21, y los descendientes de F_1 eran considerados como resistentes cuando los tests de resistencia a la raza altamente virulenta F de orobanque mostraban la presencia de los genes de resistencia predominantes. La comparación de la resistencia de las poblaciones en separación, F_2 y BC_1F_1 , con la resistencia de los componentes parentales ha confirmado la presencia de predominación notada en la generación F_1 , indicando al mismo tiempo que esta resistencia es controlada por un gen predominante. Esta predominación del gen de resistencia simplificará mucho la selección con respecto a la resistencia.

TRANSMISSION DE RÉSISTANCE À UNE RACE TRÈS AGRESSIVE F DE l'*Orobanche cumana* Wallr. DANS UNE LIGNE DE TOURNESOL DÉRIVÉE D'AMPHIPLOÏDES INTERSPECIES

RÉSUMÉ

En Espagne, les populations d'orobanche (*Orobanche cumana* Wallr.) appartenant à la nouvelle race F ne réagissent à aucun gène de résistance connu, depuis Or_1 jusqu'à Or_5 , présents dans le tournesol cultivé (*Helianthus annuus* L.) et elles se répandent rapidement. La résistance à la race F de cette herbe parasitaire a été découverte dans des espèces sauvages de tournesol et elle a été introduite dans le tournesol cultivé. Le but de cette étude était la description de la transmission des gènes de résistance dans le tournesol cultivé dérivé des espèces vivaces *H. divaricatus* et *H. grosseserratus*. Les lignes résistantes de tournesol cultivé ont été croisées avec la ligne sensible P21 et la progéniture F_1 a été considérée résistante quand les tests de résistance envers la race très agressive d'orobanche F ont montré la présence de gènes de résistance dominants. La comparaison de la résistance des populations en division, F_2 et BC_1F_1 avec celle des parents a confirmé la dominance observée dans F_1 et indiqué que la résistance était sous le contrôle d'un unique gène dominant. Cette dominance de gènes résistants simplifiera grandement la culture de plantes pour leur résistance.

