

SCREENING WILD SUNFLOWER SPECIES AND F₁ INTERSPECIFIC HYBRIDS FOR RESISTANCE TO BROOMRAPE

Terzić, S. *, Dedić, B., Atlagić, J., Jocić, S. and Tančić, S.

*Institute of Field and Vegetable Crops, Oil Crops Department,
Maksima Gorkog 30, 21000 Novi Sad, Serbia*

*Received: July 25, 2010
Accepted: November 15, 2010*

SUMMARY

Broomrape has become one of the most important threats to the cultivated sunflower. New sources of resistance genes are needed to maintain sunflower production. The objective of this research was to evaluate accessions of wild sunflower species and their F₁ interspecific hybrids with cultivated sunflower for resistance to race E.

Plant material consisted of 6 accessions of perennial species and 14 F₁ accessions between perennial species and cultivated sunflower as well as 42 accessions of annual species. Cultivated line JM8 was used as the sensitive check. Accessions were screened in the greenhouse in the season of 2008/09 and in a field trial during 2009. Plants were grown in the infested soil and evaluated for reaction at the end of the vegetation.

The cultivated check was susceptible in the greenhouse and in the field trial.

The annual species showed varying resistance with *H. annuus* and *H. argophyllus* as the most sensitive with an average of 6.6 and 8.5 broomrape plants per sunflower plant. Only 2 of the 7 *H. petiolaris* accessions were susceptible. *H. neglectus* performed well in the greenhouse with only one broomrape plant infecting one plant of the four tested accessions, but accession NEG1363 was infected in the field trial leaving NEG457 and 1183 as resistant. All accessions of perennial species except for an F₁ hybrid of DEC B and F₁ RIG 707 showed complete resistance.

New potential sources of broomrape resistance genes have been found among wild species and their interspecific hybrids.

Key words: *Helianthus annuus* L., *Orobanche cumana* Wallr., broomrape, resistance, sunflower, wild species

* Corresponding author: Phone: +381 21 4898 415; Fax: +381 21 6413 833;
e-mail: sreten.terzic@ifvcns.ns.ac.rs

INTRODUCTION

Cultivated sunflower was obtained through a selection process where seed yield and oil content were mostly emphasized (Škorić, 1988). Its quality as a cultivated plant was greatly improved in a relatively short period between 1930 and 1970 where it started from the tenth and eventually reached the second place as an oil cultivar right behind soybean. Oil content was raised from 28% (1920) to almost 50% (1955), but the disease problem was always present (Heiser, 1976). Broomrape has now become one of the most important threats to sunflower cultivation. At present, this parasitic angiosperm can be found in Eastern Europe, Spain, the Middle East and India. The yield loss depends on the severity of infection and can be very high.

Wild species of sunflower represent the source of resistance to many pathogens that attack cultivated sunflower. Because of that, it was reasonable to use wild species as a source of resistance genes in sunflower breeding programs through interspecific crosses. Genetic resistance to broomrape has been introduced to cultivated sunflower from the wild species, mainly *H. tuberosus*, into early cultivars in the former USSR (Pustovoit, 1966). The wide use of the same resistance type led to the appearance of new races (Škorić, 1988). By continuing the broomrape race nomenclature given by Vranceanu *et al.* (1980) for five physiological races (A, B, C, D and E), a total of seven races with race G as the newest one is described (Molinero-Ruiz and Melero-Vara, 2004), even though the last one could only be a population of race F (Molinero-Ruiz *et al.*, 2009).

According to Aćimović (1977), broomrape was described for the first time in Serbia in 1951, and it started causing significant damage in susceptible hybrids in the 1990s (Maširević and Medić-Pap, 2009). The first report of race E in Serbia appeared in 1996 (Mihaljčević). It was determined using a set of differential sunflower lines and broomrape seed collected in the fields in the northern part of Vojvodina. According to recent research, race E is still the most virulent race present in Serbia (Dedić *et al.*, 2009).

Because of the constant increase in selection pressure against broomrape races through introduction of new resistance genes and cultivation of resistant hybrids, new broomrape races are also occurring in shorter intervals and limit the durability of introduced resistance. The strategy of broomrape control is therefore changing, with the aim to make resistance more durable by including the use of herbicide tolerant IMI cultivars, quantitative / horizontal resistance, and by understanding the genetic structure and variability of *O. cumana* populations (Fernandez-Martinez *et al.*, 2009).

Genetic resistance is still the main component in sunflower broomrape control and resistance genes originating from wild sunflower species can have an important role in providing durable resistance. A high level of resistance to broomrape races was previously reported in wild species like *H. tuberosus*, and *H. annuus* (Fernan-

dez-Martinez *et al.*, 2008). The objective of this research is to distinguish accessions that are potential carriers of resistance gene/genes for race E by testing the wild species and their F₁ generations with cultivated sunflower. Although the resistance is more frequently found in the perennial species, the annual species are included in the test because they are much easier to cross with cultivated sunflower.

MATERIALS AND METHODS

Plant material consisted of 42 accessions of annual species (Table 1) as well as 6 accessions of perennial species and 14 F₁ accessions between perennial species and cultivated sunflower (Table 2). Cultivated line JM8 with no resistance genes was used as the sensitive check in the evaluated material. Accessions were screened in the greenhouse in the season of 2008/09 and in a field trial in 2009 for resistance to broomrape.

To promote germination the seed hull of wild species was removed (Chandler and Jan, 1985). Dehulled seeds were first kept in the dark in a growth chamber for 24 h and after the root emergence transferred to jiffy 7 pots. Seedlings were planted in jiffy 7 pots and grown in the greenhouse at 21°C (day and night), with relative humidity at around 80%, and constant light until the phase of two pairs of leaves, after which they were transferred to buckets (10 dm³) on October 15th 2008 or planted in the field for the field trial in 2009. The buckets were filled with a mixture of sand:perlite:peat in a 1:1:1 ratio which was homogeneously infested with broomrape seeds of race E at a rate of 70 mg/dm³ soil mixture. Broomrape seeds were extracted from the stalks collected in August 2007 from locations in Vojvodina where race E was previously determined. Plants were grown in the infested soil for 90 days at a photoperiod of 16/8h day/night with natural light supplemented with metal halide lamps. In the field trial, the seed material was sown at the same time as the rest of the cultivated sunflower. Reaction of plants was evaluated when they reached physiological maturity.

Plants with emerged or underground broomrape stalks were considered susceptible and those without a broomrape stalk as resistant. Severity was calculated as the number of broomrape plants per sunflower plant. Accessions that were susceptible in the greenhouse test were not included in the field trial.

RESULTS AND DISCUSSION

The cultivated check was susceptible in the greenhouse and in the field test. The annual species showed varying resistance with *H. annuus* and *H. argophyllus* as the most sensitive with an average of 6.6 and 8.5 broomrape plants per sunflower plant, respectively. All accessions of those two species were found susceptible in the greenhouse trial. First broomrape stalks emerged after 70 days in two

accessions of *H. annuus* and one of *H. niveus* and the rest emerged by day 93 of growing in the greenhouse. Broomrape emergence could be taken as additional information on broomrape resistance, especially for heterogeneous material like the accessions of wild species (Tab. 1).

Table 1: Accession identifiers and reaction of annual species to race E of *Orobanche cumana* Wallr.

Genotype/ wild species	IFVCNS population	PI number	Greenhouse		Field trial	
			Intensity of attack	Broomrape emergence (DAS [§])	Intensity of attack	Broomrape emergence (DAS)
<i>H. debilis</i>	1134	PI 468678	0	-	-	-
<i>H. debilis</i>	1136	PI 468680	0	-	2.4	79
<i>H. debilis</i>	1290	PI 468687	8	*	-	-
<i>H. debilis</i>	1675	PI 468692	0	-	-	-
<i>H. debilis</i>	1810	PI 494583	3.5	93	-	-
<i>H. debilis</i>	1848	PI 494589	0	-	3.5	97
<i>H. annuus</i>	1970	PI 531015	1.5	86	-	-
<i>H. annuus</i>	2034	PI 547165	8	74	-	-
<i>H. annuus</i>	2104	PI 586809	11.5	74	-	-
<i>H. annuus</i>	2150	PI 586839	9	70	-	-
<i>H. annuus</i>	2191	PI 586866	0.5	-	-	-
<i>H. annuus</i>	2206	PI 586877	3.5	70	-	-
<i>H. annuus</i>	2220	PI 586883	12	74	-	-
<i>H. argophyllus</i>	1317	PI 468649	4	93	-	-
<i>H. argophyllus</i>	1575	PI 468651	6.5	93	-	-
<i>H. argophyllus</i>	1807	PI 494573	15	86	-	-
<i>H. petiolaris</i>	338	PI 435803	0	-	0	-
<i>H. petiolaris</i>	722	PI 435831	0	-	0	-
<i>H. petiolaris</i>	815	PI 435845	0	-	2.1	93
<i>H. petiolaris</i>	1383	PI 468811	1.5	80	-	-
<i>H. petiolaris</i>	1910	PI 503232	0	-	0	-
<i>H. petiolaris</i>	2146	PI 586918	0	-	0	-
<i>H. petiolaris</i>	2208	PI 586931	0	-	0	-
<i>H. praecox</i>	1142	PI 468851	0	-	7.1	79
<i>H. praecox</i>	1151	PI 468854	0.5	*	-	-
<i>H. praecox</i>	1801	PI 494606	1.5	*	1.8	79
<i>H. praecox</i>	1333	PI 468865	0	-	5.2	86
<i>H. praecox</i>	1341	PI 468849	0	-	0	-
<i>H. praecox</i>	1168	PI 468847	0	-	7	90
<i>H. praecox</i>	1821	PI 494610	0,5	*	0	-
<i>H. niveus</i>	1403	PI 468784	0	-	3.9	86
<i>H. niveus</i>	1452	PI 468788	6,5	70	-	-
<i>H. neglectus</i>	1181	PI 468765	0,5	*	-	-
<i>H. neglectus</i>	1363	PI 468775	0	-	5.6	86
<i>H. neglectus</i>	457	PI 435761	0	-	0	-
<i>H. neglectus</i>	1183	PI 468767	0	-	0	-
JM 8	Cultivated		2,3	86	7.8	79

* All broomrape stalks were underground at the moment of evaluation

§ DAS – Days after sowing

Similar findings in regard to the resistance of *H. annuus* and *H. argophyllus* were presented by Fernandez-Martinez *et al.* (2008), who also described the species *H. petiolaris* and *H. praecox* as 100% susceptible. *H. petiolaris* performed differently in this trial with only 2 out of the 7 accessions susceptible. Accession PET1383 had 1.5 broomrape plants per sunflower in the greenhouse and was not tested in the field and accession PET815 was not infected in the greenhouse but had 2.1 broomrape plants per sunflower of infected plants in the field trial. Different resistance could be simply a result of different accessions that were used or different broomrape race composition.

H. debilis had two resistant accessions (DEB1134 and DEB 1675) out of the six tested. The rest were infected either in the greenhouse (DEB1290, 1810) or in the field trial (DEB1136, 1848). *H. neglectus* performed very well in the greenhouse, with only one broomrape plant infecting one plant of the four tested accessions, but accession NEG1363 was infected in the field trial leaving NEG457 and 1183 as resistant.

Table 2: Accession identifiers and reaction of perennial species and their F₁ hybrids with cultivated sunflower to race E of *Orobanche cumana* Wallr.

Genotype /wild species	IFVCNS population	(Wild parent) PI number	Greenhouse		Field trial	
			Intensity of attack	Broomrape emergence (DAS [§])	Intensity of attack	Broomrape emergence (DAS)
F ₁ <i>H. rigidus</i>	1693	-	0	-	0	-
F ₁ <i>H. rigidus</i>	1696	-	0	-	0	-
F ₁ <i>H. rigidus</i>	707	-	0	-	2.1	79
F ₁ <i>H. tuberosus</i>	1698	-	0	-	0	-
F ₁ <i>H. tuberosus</i>	1701	-	0	-	0	-
F ₁ <i>H. tuberosus</i>	6	-	0	-	0	-
F ₁ <i>H. strumosus</i>	1927	PI 503249	0	-	0	-
F ₁ <i>H. strumosus</i>	1623	PI 468894	0	-	0	-
F ₁ <i>H. divaricatus</i>	2085	PI 547174	0	-	0	-
F ₁ <i>H. hirsutus</i>	1536	PI 468738	0	-	0	-
F ₁ <i>H. eggertii</i>	1626	PI 468712	0	-	0	-
F ₁ <i>H. decapetalus</i>	B	-	0.5	*	0	-
F ₁ <i>H. resinosus</i>	1545	PI 468879	0	-	0	-
F ₁ <i>H. laevigatus</i>	1618	PI 468740	0	-	0	-
<i>H. tuberosus</i>	675	PI 468829	0	-	-	-
<i>H. tuberosus</i>	26	-	0	-	-	-
<i>H. grossesseratus</i>	56	-	0	-	-	-
<i>H. mollis</i>	1530	PI 468760	0	-	-	-
<i>H. nuttallii</i>	239	PI 435779	0	-	-	-
<i>H. tuberosus</i>	CG 56	-	0	-	-	-
JM 8	Cultivated		2,3	86	7.8	79

* All broomrape stalks were underground at the moment of evaluation

§ DAS – Days after sowing

All evaluated accessions of perennial species showed complete resistance. Similar results for perennial species was obtained by other authors (Fernandez-Martinez *et al.*, 2000) which is why they were not included in the field trial to limit the possibility of unwanted propagation. It is worth noting that only two out of the 14 tested F₁ crosses between cultivated sunflower and perennial species were susceptible. An F₁ hybrid with DEC B accession reached physiological maturity in the greenhouse after approximately 90 days with an attack intensity of only one underground broomrape stalk per sunflower. An F₁ hybrid with RIG 707 was not infected with broomrape in the greenhouse, but in the field trial it had 2.1 broomrape plants per sunflower (Table 2). It has been shown that the wild species and especially perennials have a great potential in providing resistance for the future broomrape races. Although the resistance was higher in perennial species and their F₁ progenies, resistant plants found among the annual species are very important in regard to the short-term breeding programs. It is always better to use the resistance genes from the gene pool of cultivated sunflower, but in some case, like with the newest races of broomrape, it is fortunate that the interspecific crosses are possible inside the genus *Helianthus*.

REFERENCES

- Aćimović, M., 1977. Distribution of important sunflower diseases in Yugoslavia. Periodical of Scientific research on field and vegetable crops, Faculty of Agriculture, Institute of Field and Vegetable Crops, Novi Sad, Yugoslavia 1: 1-16.
- Chandler, J.M., Jan, C.C., 1985. Comparations of germination techniques for wild *Helianthus* seeds. Crop Science 25: 356-359.
- Dedić, B., Lačok, N., Tančić, S., Dušanić, N., Jocić, S., 2009. Current status of broomrape (*Orobanche cumana* Wallr.) in Serbia. Helia 32(51): 135-140.
- Fernandez-Martinez, J.M., Melero-Vara, J., Munoz-Ruz, J., Ruso, J., Dominguez, J., 2000. Selection of wild and cultivated sunflower for resistance to a new broomrape race that overcomes resistance of the *Or₅* gene. Crop Science 40(2): 550-555.
- Fernandez-Martinez, J.M., Dominguez, J., Perez-Vich, B., Velasco, L., 2008. Update on breeding for resistance to sunflower broomrape. Helia 31(48): 73-84.
- Fernandez-Martinez, J.M., Dominguez, J., Perez-Vich, B., Velasco, L., 2009. Current research strategies for sunflower broomrape control in Spain. Helia 32(51): 47-56.
- Maširević, S., Medić-Pap, S., 2009. Broomrape in Serbia from its occurrence till today. Helia 32(51): 91-100.
- Molinero-Ruiz, M.L., Melero-Vara, J.M., 2004. Virulence and agresiveness of sunflower broomrape (*Orobanche cumana*) populations overcoming the *Or₅* gene. In: Seiler, G.J. [ed.], Proc. 16th Intl. Sunflower Conf., Fargo, ND, USA, 29 August-4 September 2004. Intl. Sunflower Assoc., Paris, France 1: 165-169.
- Molinero-Ruiz, M.L., Garcia-Ruiz, R., Melero-Vara, J.M., Dominguez, J., 2009. *Orobanche cumana* race F: performance of resistant sunflower hybrids and aggressiveness of populations of the parasitic weed. Weed Research 49: 469-478.
- Pustovoit, V.S., 1966. Selection, seed culture and some agro technical problems of sunflower. [Translated from Russian in 1976 by INSDC], Indian National Scientific Documentation Centre, Delhi, India.
- Škorić, D., 1988. Sunflower breeding. Uljarstvo 25(1): 1-90.
- Vranceanu, A.V., Tudor, V.A., Stoenescu, F.M., Pirvu, N., 1980. Virulence groups of *Orobanche cumana* Wallr. differential hosts and resistance sources and genes in sunflower. In: Proc. 9th Intl. Sunflower Conf., Torremolinos, Spain, 8-13 July 1980. Intl. Sunflower Assoc., Paris, France 1: 74-80.