DILEMMAS ABOUT NEW SUNFLOWER BROOMRAPE RACES (Orobanche cumana Wallr.)

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The genus Orobanche

The genus Orobanche contains over 100 species

Broomrape species have been reported in a total of 58 countries

Orobanche cumana Wallr = Orobanche cernua Loefl.



Orobanche on different plant species...













Orobanche on different plant species...











Orobanche reticulata ssp. reticulata







Orobanche on sunflower + ? ! ...



Orobanche cummana Wallr. on sunflower



Orobanche cumm on sunflov







Orobanche on sunflower + ? ! ...







There is permanent combat between breeders and geneticists on one side, and Orobanche on the other, and the WINNER is often found at different sides;

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SEEDS AND GERMINATION

Seed and germination

The fruit is a capsule, the seeds are brown, 0.2-0.4 мм in size;

Flower number per plant: 30-67;

Seed number per flower: 2,500-10,500; Absolute seed weight: 0.001 g;

Seed number per plant: > 87,000;

Number of seeds: >44,000/g;

Broomrape is transported by wind and vehicles (combines, tractors, trucks...)

mm

SEEDS AND GERMINATION

Broomrape seeds may lie dormant in the soil for more than 20 years and still remain viable;

The seeds germinate in moist soil at a temperature of 20-25°C;

The seedlings take the form of a thin strand – the growth shows positive gravitropism;

If the seedling does not come into contact with the root of the host plant, it will die relatively quickly;

Germinability is affected by the pH of the environment as well as by the root exudates of the host (germination stimulants);

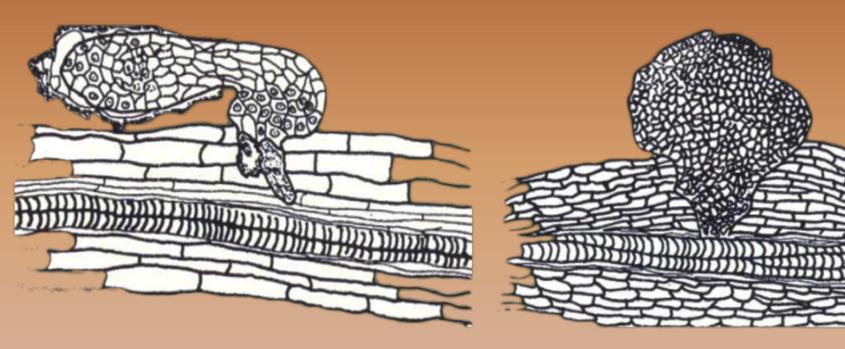
Range of physiological races of *Orobanche cumana Wallr*. at Constanta, Tulcea, and Braila (Romania) (Vranceanu *et al*. (1980), Pacureanu *et al*. (1998))

	Resistance gene	Natural infestation - Constanta, 1997					Inoculation, Fundulea, 1997						
Differential line		Sunflower line response to broomrape race						Sunflower line response to broomrape race					
		Α	B	С	D	Ε	F	Α	B	С	D	Ε	F
LC-1093	0r6	R	R	R	R	R	R	R	R	R	R	R	R
P-1380-2	Or5	R	R	R	R	R	S	R	R	R	R	R	S
S-1358	Or4	R	R	R	R	S	S	R	R	R	R	S	S
Record	Or3	R	R	R	S	S	S	R	R	R	S	S	S
Zhdanov	Or2	R	R	S	S	S	S	R	R	S	S	S	S
Kruglik A-41	Or1	R	S	S	S	S	S	R	S	S	S	S	S
AD-66		S	S	S	S	S	S	S	S	S	S	S	S

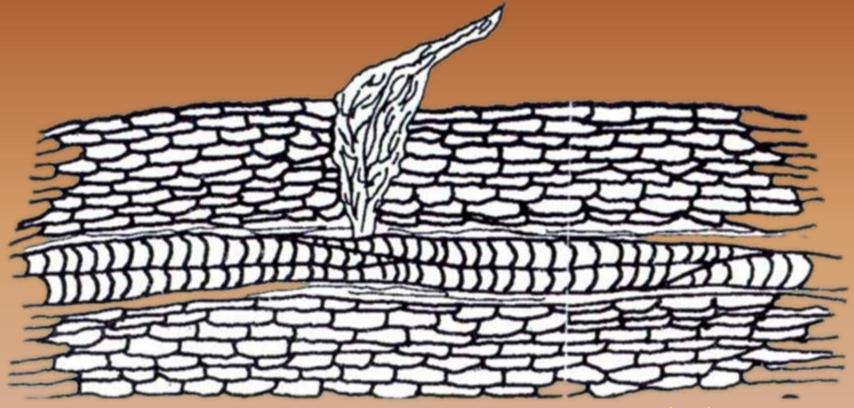


1. Germination of broomrape seeds, the penetration of the root, and the formation of a primary haustorium during the infection of resistant and susceptible forms of sunflower (8 days after germination)

2. The haustorium penetrating the central cylinder of the radicle (xylem) in a susceptible sunflower genotype (14 days after germination)



3. Broomrape germ dying in a resistant sunflower genotype



Panchenko, 1975

There are different mechanisms of resistance to broomrape:

MECHANISMISIOF RESISTANCE TO

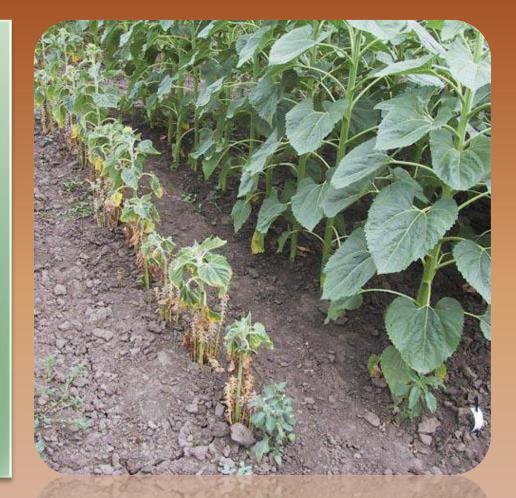
Orobanche cumana Walk



SOME OF THE PARAMETERS OF SUNFLOWER RESISTANCE TO Orobanche cumana Wallr.

According to Richter (1924) (as cited by Morozov, 1947):

- Susceptible sunflower plants have low pH values of the root system (acidity);
- Two physiological thresholds: 1. In acid soils, broomrape seeds germinate easily; 2. In alkaline ones, susceptible sunflower genotypes become "resistant" and no broomrape infestation occurs.



SOME OF THE PARAMETERS OF SUNFLOWER RESISTANCE TO Orobanche cumana Wallr.

According to Suhorukov (1930) (as cited by Morozov (1947):

- Increased soil acidity leads to an increase in peroxidase activity, resulting in sunflower nonresistance to the pathogen;
- Ukrajinski (1947) notes that broomrape resistance is determined by the specific responses of the sunflower root system;
- The author proved that resistant sunflower genotypes have a low pH at the start of the season and a high pH at the season's end.



SCREENING METHODS

Planting the breeding material on naturally infested plots Evaluation of field resistance - introduction of broomrape seeds at sunflower planting

Resistance evaluation in the greenhouse – planting in pots (sunflower seeds + broomrape seeds)







Rapid evaluation of resistance in the greenhouse – planting on tables (soil + broomrape seeds + sunflower seeds)



According to Panchenko





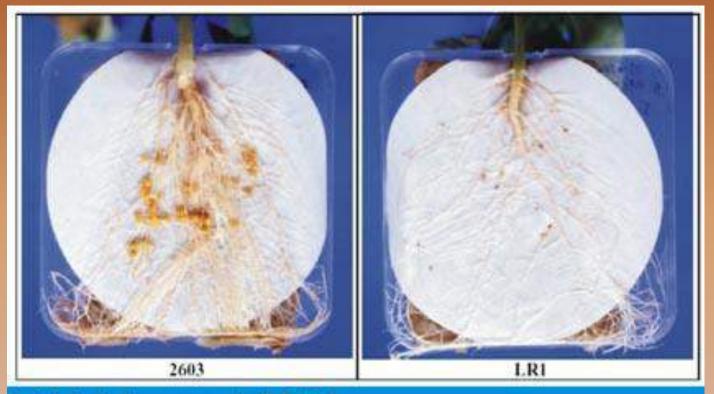


Rapid evaluation of resistance in the greenhouse - planting in test tubes

> substrate + broomrape seeds + sunflower seeds

> > (Grezes-Besset) Improved by Mihaljcevic

Hydroponic co-culture of susceptible (2603) and resistant (LR1) sunflower parasitized by broomrape, 21 days after infestation.



Patricia Labrousse et al. 2004.

Physiological features of possible regulation of sunflower resistance to broomrape (*Orobanche cumana* Wallr.) _{Kyrychenko et al.} (2018)

1. Physiological aspects of sunflower resistance to broomrape (*Orobanche cumana* Wallr.)
2. Influence of broomrape affection on morphological and physiological parameters of sunflower plants
2.1. Total content of phenolic compounds in broomrape (*O. cumana* Wallr)-infected sunflower plants.
2.2. Activity of oxidizing/reducing enzymes in different genotypes of sunflower infected by angiosperm parasite.

Physiological features of possible regulation of sunflower resistance to broomrape (*Orobanche cumana* Wallr.) Kyrychenko *et al.* (2018)

• Broomrape infection inhibits the growth of susceptible sunflower plants, however, it hardly affects morphological parameters of resistant sunflower genotypes. It was revealed that resistant and susceptible genotypes differed by phenol levels in plants under the influence of Orobanche cumana Wallr. The values of resistant accessions were higher than those of susceptible ones. It was determined that infection sunflower plants by angiosperm parasite considerably affected enzyme activities.

Physiological features of possible regulation of sunflower resistance to broomrape (Orobanche cumana Wallr.) _{Kyrychenko et al.} (2018)

 The most distinct patterns were observed on day 14 after infection, which indicates the feasibility of conducting research during this period. It was shown that the activities of enzymes (polyphenol oxidase, peroxidase and catalase) significantly increased, except those in the linesusceptibility standard, the values of which remained almost unchanged or substantially reduced after broomrape infection.

 Basing on our data, we developed a biochemical expressmethod for assessing sunflower genotypes for resistance to *Orobanche cumana* Wallr. The method consists in determination of catalase activity in green plant material.



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BREEDING METHODS:

Conventional methods

Induced methods

MAS (Marker-Assisted Selection)

Somatic hybridization

Genetic transformation (GMO)



SOURCES OF RESISTANCE:

Varietal populations

Landraces

Inbred lines

Hybrids resistant to broomrape

Interspecific hybrids (cultivated x wild)

Wild species of the genus *Helianthus*



Wild sunflower species containing genes for resistance to *Orobanche cumana* Wallr.

Helianthus tuberosus;	H.ciliaris;				
H.smithii;	H.resinosus;				
H.pauciflorus;	H.laevigatus;				
H.nuttallii ssp. nuttallii;	H.gigantheus;				
H.pumilus;	H.grosseserratus;				
H.maximiliani;	H.agrestis;				
H.hirsutus;	H.anomalus;				
H.eggertii;	H.debilis ssp.cucumerifolius;				
H.divaricatus;	H.exilis;				
H.decapetalus	H.rigidus				

Table 1: The degree of broomrape (Orobanche cumana Wallr.)resistance in different sunflower genotypes at various Romanianlocalities in 2014 (Number of broomrape plants/main plot)

No	Comotomo	Locality						
No.	Genotype	Cuza Voda	Crucea-Stupina	Valea Canepii	Tulcea-Agighiol			
1	LC-1093 (rase F)	542	154	1.136	267			
2	Performer	1.532	437	3.397	432			
3	LG-5661	48	28	40	10			
4	PR64LE20	69	58	129	43			
5	No. 97	0	46	70	18			
6	No. 144	0	72	4	9			
7	No. 185	0	3	1	4			
8	No. 194	0	32	130	25			
9	No. 242	0	64	17	11			
10	No. 341	0	30	123	35			
11	No. 378	0	2	1	0			
12	No. 388	0	110	63	10			
13	No. 367	38	0	1	2			
14	No. 143	-	38	0	14			
15	No. 9	60	100	246	0			
16	No. 188	20	8	1	0			
17	No. 190	2	36	1	0			
18	No. 191	3	33	5	0			
19	No. 219	150	53	280	0			
20	No. 274	40	40	28	0			
21	No. 302	50	7	45	0			
22	No. 348	1	9	3	0			
23	No. 365	3	18	15	0			
24	No. 381	8	23	4	0			
25	No. 383	1	18	1	0			
26	No. 384	2	7	2	0			
27	No. 386	0	12	15	0			

Table 2: The degree of broomrape (Orobanche cumana Wallr.)resistance in different sunflower genotypes at various Romanianlocalities in 2015 (Number of broomrape plants/plot)

		Localities									
		Ciresu-Braila		Iazu-Ialomita		Stupina-Constanta		Topolog-Tulcea		Viziru-Braila	
No.	No. Genotype	Sunflower plants/plot	Broomrape plants/plot	Sunflower plants /plot	Broomrape plants /plot						
1	Performer	89	679	48	97	39	102	46	169	45	204
2	LC-1093	47	41	43	8	32	41	43	51	46	19
3	LG-5542	45	0	-	-	38	5	46	6	46	0
4	LG-5631	46	12	-	-	-	-	-	-	-	-
5	PR64LE20	48	16	44	0	41	18	44	26	44	0
6	Hy-1	128	887	95	136	47	68	95	113	105	38
7	Hy-2	120	798	105	87	55	95	95	233	94	98
8	Hy-7	126	0	95	0	44	0	96	0	141	0
9	Hy-9	124	0	106	1	55	11	82	18	100	8
10	Ну-10	132	2	94	0	46	16	93	59	95	30

Table 3: Evaluation of broomrape (*Orobanche cumana* Wallr.) resistance in different sunflower genotypes at the infested plot: VNIIMK – Don trial station, Rostov on Don in 2017

No.	Genotype	Percentage of infested sunfloer plants/main plot	Degree of infestation
1	Donskoy-22	100	
2	Bella	30	3,0
3	LG-5580	23	1.0 - 1.2
4	PR66LE25	19.4	2.6
5	Hy-7 (F ₁)	17.1	1.1
6	F ₂ (Hy-7)	35.1	2.0



 However, speaking of the new races G and H, we might not be completely sure. The question is whether the same mutation can occur in broomrape populations, in various countries during the same year? Certain differences which have not yet been studied in detail surely exist.



The question regarding the degree of virulence in the new broomrape populations remains unanswered.
 Significant differences have best been illustrated by Pacureanu-Joita *et al.* (2012), stating that the most aggressive populations were found in Moldova, Romania, Russia, and Turkey.

• New studies worth mentioning are the ones carried out by Duca *et al.* (2017), who performed molecular characterization of 39 broomrape populations from Moldova using SSR markers.

 Based on the obtained results, the authors concluded that some populations from the Southern region (especially Carabetovca, Alexanderfeld, Stefan-Voda, and Slobozia Mare) have shown major differences in the obtained profiles and presented a high degree of genetic variability. The study revealed genetic diversity of *O. cumana* populations and contributed to research by providing useful information on this economically important pathogen.



• In conclusion, the results obtained in this research partly differ from the results obtained by Rişnoveanu et al. (2016), which was to be expected since the trials were carried out at various localities (plots). However, the results concur with authors' conclusion, confirming identification of new races of Orobanche previously undetected at the trial localities.

BROOMRAPE CONTROL BY HERBICIDES

Herbicide group:

• imidazolinones (imazethapyr, imazamax, imazapyr)

Sources of resistance:

- Wild Helianthus annuus L.
- (discovered by Al- Khatib *et al.,* 1998 a crop of soybean)
- Genes incorporated into elite lines of cultivated sunflower by backcrossing (BC) + screening for resistance
- Resistance controlled by two partially dominant genes (*Imr1; Imr2*), *Imr2* modifier gene.

- Novi Sad sunflower team was among the first to develop commercial IMI-resistant hybrids
- Sala et al. (2008) identified a new source of resistance using induced mutations
- (EMS ethyl methanesulfanate, 0.25% solution)
- New IMI-resistance gene: CLHA PLUS (partial dominance)
- CLHA–PLUS differs from *Imri*
- Excellent weed control + control of *Orobanche*!



Resistance in wild sunflower...

At the beginning of the 21st century, public resistance sources were found in the wild sunflower *Helianthus annuus* L. discovered in Kansas (USA). Resistance source named CHLA- PLUS, obtained by Sala (2008) through induced mutations, has lately been used by the company BASF.

It has been shown experimentally that the gene CHLA-PLUS a higher degree of IMI-resistance than the gene Imr, Imr₂.
 Breeding centers wishing to use the CHLA-PLUS gene for breeding purposes have to sign a contract on its use with the company BASF. At the same time, BASF provides a protocol for screening for resistance at the molecular level (CLEARFIELD[®] Protocol SF₃0).

Resistance to broomrape...

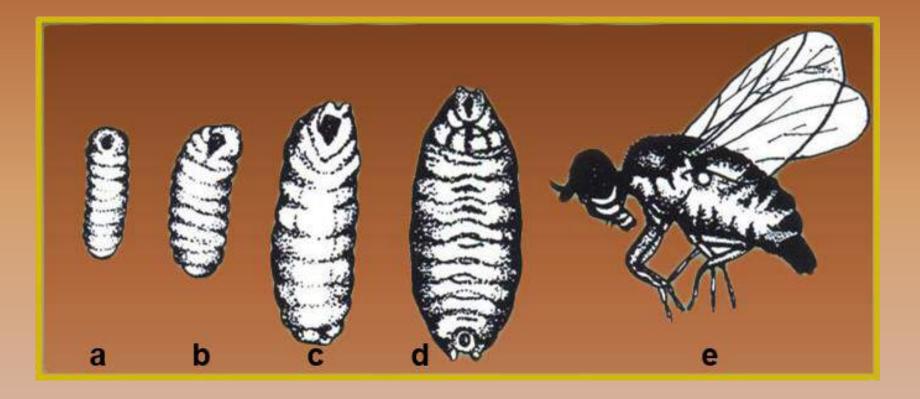
- The results of Gorbachenko *et al.* (2018) suggest the possibility of developing new cultivars of sunflower resistant to the new variability of broomrape populations.
- This method of testing broomrape resistance has its drawbacks due to the differences in broomrape distribution at the testing plots. Also, broomrape expression depends on several factors, such as soil type, distribution and amount of rainfall during the vegetation period, temperature, pH value, and so forth.

• The results of Gorbachenko *et al.* (2018) offer the possibility of developing sunflower hybrids resistant to the new variability of broomrape populations which cannot be controlled by genes for race H.



Biological control of Orobanche

INSECT: *Phytomyza orobanchia Kalt.*



WHAT SHOULD BE DONE NEXT?

An international project of global magnitude needs to be established;

Because of quarantine limitations, an organization should be designated to store seeds of Orobanche cumana Wallr. from all over the world and to make them available for use on global level;

An organization should also be identified that would be willing to investigate in detail the genetic variability of broomrape on the molecular level;

Genotypes of cultivated sunflower and wild *Helianthus* should be tested using the total variability of *Orobanche*;

WHAT SHOULD BE DONE NEXT?

- All all

Wild sunflower species possessing complete resistance to total Orobanche variability should be used for incorporating the resistance genes into elite lines of cultivated sunflower;

It should be established which organizations in the world would be interested in taking part in such a project (public + private sector);

Issues of intellectual property rights and the right to use the new genetic variability should be settled in advance;

Efforts should be made to identify those in whose interest it would be to provide funding for a global project of this kind;

Conclusions

Based on the studies in sunflower genetics and breeding for broomrape resistance in the Black Sea area, especially in Romania and Russia, the following conclusions can be made:

- Methods have been developed for the evaluation of resistance to Orobanche;
- Conventional methods and MAS (Marker Assisted Selection) have been developed in breeding for resistance to Orobanche;



Conclusions

• In Romania, new virulent population (races) of broomrape have emerged, especially in the regions of Braila, Tulcea, and Constanta. At certain localities, new variability of broomrape populations has been observed, which cannot be controlled by genes for race H;



 Likewise, at the trial station of VNIIMK – Don Trial Station in Russia, a new virulent broomrape population emerged, which cannot be controlled by genes for race H. Luckily, sunflower breeders have developed new genotypes resistant to the new variability of broomrape populations;

Conclusions

- There is permanent change in populations of *Orobanche cumana* Wallr., thus hindering the success of breeding for resistance;
- Sources of broomrape resistance genes are found in certain wild species of *Helianthus*, which are transferred after detection to the elite lines of cultivated sunflower using interspecies hybridization;



 Besides control through genes for broomrape resistance, Orobanche can successfully be controlled by the development of IMI-resistance hybrids.

Thank you for your attention!

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