Insecticide residues cross-contamination of oilseeds during storage

Sylvie Dauguet, Jean-Philippe Loison

CETIOM, Service Transformation et Valorisation des graines oléagineuses, 11 rue Monge Parc Industriel 33600 PESSAC, France, E-mail: dauguet@cetiom.fr

ABSTRACT

Pesticide residues are found in oilseeds and crude oils: they are mainly organophosphate insecticides (pirimiphos-methyl, dichlorvos, malathion) used in empty storage facilities and for application to stored cereal grains. Even if pests are found in stored oilseeds, French regulations do not permit the use of these insecticides on stored oilseeds, as they have an affinity for these lipophilic substances. These residues arise from cross-contamination from storage bins and facilities, and not from illegal use. This uptake of insecticide residues from their storage environment by oilseeds can lead to levels that exceed regulatory limits. An investigation of 11 grain storage companies allowed us to follow the course of 27 sunflower seeds batches, from reception at the storage facilities to outloading. Samples from each of these batches, made at outloading, were analysed by ITERG (French Technical Institute for Oil Industry), looking for insecticide residues. Traceability of sunflower seeds established by storers allowed us to identify crosscontamination sources. Substances discovered were dichlorvos, pirimiphos-methyl and malathion (and one case with chlorpyriphos-methyl). Pirimiphos-methyl was most commonly detected, but most cases of non-accordance with regulatory levels were caused by dichlorvos and malathion. Main crosscontamination hazard resulted from treatment of cereals at outloading, just before sunflower seeds were outloaded, especially when these cereal treatments were frequent on that elevator. Other situations led to cross-contamination, but generally at lower levels: outloading of sunflower seeds after outloading of treated cereal, sunflower seeds stored in bin that contained previously treated cereals, empty bins and handling equipment treated before receipt of sunflower seeds.

Key words: cross-contamination – insecticide – oilseeds – pesticide residues – storage.

INTRODUCTION

Post-harvest insecticide residues are frequently found on oilseeds at low levels, although no insecticide is allowed to be applied directly to oilseeds during storage. Consequently, maximum residue levels (MRLs) allowed by European regulation are very low (mostly at the lower limit of analytical determination): 0.01 mg/kg for dichlorvos (still authorised during this study, but forbidden now), and 0.05 mg/kg for pirimiphos-methyl. No MRL exists for malathion, so it should not be found beyond the analytical limit of quantification. These insecticide treatments are authorised on stored cereals and corn as a grain protectant, and on empty storage and handling equipment as a control agent for residual insect populations in empty granaries. Dichlorvos, malathion and pirimiphos-methyl were the substances most employed during this study (storage season 2006-2007, regulations changed later).

So, we can hypothesise that cross-contamination phenomena can exist, between these various kinds of seeds, cereals and oilseeds, sharing the same grain handling and storage system. This phenomenon has already been demonstrated in Canada on rapeseed (Watter and Nowicki, 1982, 1985; White, 1983), when empty bins were treated with organophosphorous insecticides (bromophos, malathion, fenitrothion). Canadian storers were warned that treating bins before storing rapeseed could lead to residues above the maximum allowable limits.

Uptake of pirimiphos-methyl by a single-layer of rapeseed or wheat on galvanized-steel surfaces was demonstrated in a laboratory study (Dauguet et al., 2006, 2007). It was shown that, for small bins (less than 50 tons), it could lead to residue quantities above regulatory limits. But in big elevators, insecticide uptakes by seeds can also occur at other stages: conveyor belts, handling of oilseeds after cereals had been treated in the same circuits, outloading bin, etc. Therefore we cannot rule out either risk for grain storage companies.

In order to improve our knowledge about this post-harvest insecticide cross-contamination, especially in big elevators, an investigation was carried out with the collaboration of several French grain storage companies. Real cases were observed, with an accurate traceability of sunflower seeds lots all along their route inside storage facilities (from receipt to outloading) to find where the insecticides were taken up by the oilseeds. Results presented in this article were obtained in the first year of the investigation, concentrating on sunflower seeds during the storage season 2006-2007. This investigation will continue on rapeseed during the next storage season.

MATERIALS AND METHODS

The process adopted for this survey was:

- Identifying with storage operators sunflower lots that could be "traced" (recording of each step from receipt to outloading):11 grain storage companies agreed to collaborate, and allowed us to follow 27 sunflower seed bins. These companies were situated throughout the French sunflower crop area.

- Making a mean sample from each batch representative of sunflower seeds arriving at the storage facilities ("first sample") and preserving it. These samples were analysed only when residues were found in the final sample, in order to know if contamination occurred before receipt of grain. These "first samples" were analysed for 4 batches of seeds.

- Making a mean sample representative of outloaded sunflower seeds, "final sample", when the traced lot is commercialized (from one to eight months after harvesting). These "final samples" were always analysed. In one case, we had 2 samples for one sunflower batch, so that we analysed 28 final samples. The sampling method used was based on a standard method (moving seeds, for contaminant with heterogeneous distribution determination, prEN ISO 24333:2006): 25 elementary samples for 500 tons evenly distributed during the outloading (one elementary sample each 20 tons). This method was usually well observed by the commercial operators.

- Filling in a questionnaire called "traceability" which recorded each step from receipt to outloading. Operators had to indicate if treatments were applied on empty bins or handling equipment, or if cereals were treated at their receipt or outloading and if these cereals used the same conveyer circuit inside the storage facilities just before the sunflower seeds.

- Determination of insecticide residues in all the "final samples": the analytical laboratory of ITERG conducted these determinations, using the "common method" developed three years ago by a group of about twenty French laboratories (public and private) coordinated by CETIOM and ITERG: Soxhlet extraction of oil with hexane (NF EN ISO 659) was followed by analysis of organophosphorous residues by gas chromatography with NPD detection.

RESULTS

Twenty-eight samples were analyzed (Table 1). The insecticides used on cereals and for storage facilities treatment were detected: dichlorvos, pirimiphos-methyl, malathion and chlorpyriphos-methyl (only one case). Most commonly detected substance was pirimiphos-methyl: detected in 61% of samples and quantified in 39% of samples. But, malathion and dichlorvos were more frequently above MRL: 21% and 18% of cases, respectively.

On the whole, final samples were slightly contaminated as half of them contained less than 12 μ g/kg of insecticide residues (sum of residue median), and 90% of them contained less than 120 μ g/kg (sum of residues 9th decile).

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	LQ	MRL	Mean	Median	Standard deviation	9th decile	Maxi	% samples≥ LD	$samples \ge LQ$	% samples > MRL
Dichlorvos	10	10	21	0	79	27	422	32%	29%	21%
Pirimiphos-methyl	10	50	19	5	55	29	295	61%	39%	4%
Chlorpyriphos méthyl	10	50	0	0			10	4%	4%	0%
Malathion	10	-	8	0	25	17	125	18%	18%	18%
Sum of residues			48	12	102	120	427			

Table 1. Analytical results (expressed in $\mu g/kg$) on the 28 final samples¹

^TLQ: limit of quantification; LD: limit of detection; MRL: maximum residues limits in sunflower seeds; Sum of residues: a value of 5 μ g/kg is given when a substance is detected but below the limit of quantification, and zero value if under the limit of detection.

Analytical results for each substance (Fig. 1, Fig. 2, Fig. 3): Pirimiphos-methyl - Only one sample had a very high level (T12: 295 μ g/kg). The other samples were always below the MRL: 4 between 20 and 50 μ g/kg, 12 between 10 and 20 μ g/kg, and 11 below the limit of quantification; Dichlorvos - Only one sample had a very high level (T9: 422 μ g/kg). Three other samples were between 20 and 50 μ g/kg.

Five samples were near the MRL or below it. It was not detected in 19 samples; Malathion - Only one sample had a high level (T28: 125 μ g/kg). Four samples were between 10 and 50 μ g/kg, 12 between 10 and 20 μ g/kg. It was not detected in 23 samples.



Fig. 1. Individual analytical results for pirimiphos-methyl



Fig. 2. Individual analytical results for dichlorvos



Fig. 3. Individual analytical results for malathione

In order to explain the contamination, when large residues were detected, the "first samples" (taken at receipt of the sunflower seeds at the storage facilities) were analyzed for the batches T4, T12, T19 and T28. The insecticide residues on each "first sample" were too low to explain the residues found at the end, in the "final samples". So, the explanation had to be found in the route of the sunflower seeds inside the elevator.

Four cases leading to cross-contamination were identified:

- K1: treatment of cereals at outloading, just before outloading of sunflower seeds
- K2: outloading of cereals, treated at their receipt, just before outloading of sunflower seeds
- K3: storage of treated cereals in the same bin just before storage of sunflower seeds
- K4: treatment of empty bin and of handling equipment before receiving sunflower seeds



Fig. 4. Distribution of the four cases (K1, K2, K3, K4) for each sunflower lot, and sum of insecticide residues (µg/kg) for each sunflower final sample

It would seem that the biggest cross-contamination occurred with the case K1. Studying the circumstances of K1, the risk was higher when treatment of cereals at outloading was frequent or systematic. In the worst case (T9), the sunflower seeds batch was transported by lorry over a period of two weeks. Cereals were also sent during this period, and treated before outloading, and using the same outloading circuit as the sunflower (conveying belts, outloading bin). The other cases, K2, K3, K4, may also lead to a slighter cross-contamination. For one lot, there were two cases of a cross-contamination risk, and this could only have worsened the contamination.

CONCLUSIONS

Our study in real situations showed that cross-contamination of oilseeds by post-harvest insecticide residues exists, and can sometimes lead to residues above the regulatory limits.

The highest risk of contamination appeared when cereals were systematically treated at outloading, just before outloading of oilseeds, using the same conveyor circuits. The other identified cases may also lead to a slighter contamination. But, silo operators should concentrate on the accumulation of several risky cases, which could worsen the contamination.

Other sources of insecticide residues can occur in storage facilities, but we could not check them in this investigation. They include leak of insecticide from the application equipment, use of sampling equipment contaminated by pesticides. Another situation that we did not meet in our study was cross-contamination or accidental treatment on-farm of oilseeds subsequently delivered to a commercial store.

This investigation will be continued with rapeseed. This will allow us to check if rapeseed can be affected by the same cross-contamination as sunflower. The new work will be carried out in the new regulatory context in which dichlorvos and malathion are forbidden for cereal treatment. Thus, storage operators will certainly have new grain protection strategies, which could lead to a different cross-contamination risk for oilseeds.

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REFERENCES

- Dauguet, S., F. Lacoste, B. Ticot, J.P. Loison, J. Evrard, B. Bouchtane, and B. Soulet. 2006. La filière oléagineuse se mobilise autour de la problématique des résidus d'insecticides. OCL 13(6):373-377.
- Dauguet, S., J. Evrard, J. Fritsch, and J.P. Loison. 2007. Accumulation of pesticide residues in oil during the storage of rapeseed. 12th International Rapeseed Congress, Wuhan Hubei, China, March 26-30.
- Watters, F.L., and T.W. Nowicki. 1982. Uptake of bromophos by stored rapeseed. J. Econ. Entomol. 75:261-264.
- White, N.D.G, and T.W. Nowicki. 1985. Effects of temperature and duration of storage on the degradation of malathion residues in dry rapeseed. J. Stored Prod. Res. 21:111-114.
- White, N.D.G, T.W. Nowicki, and F.L. Watters. 1983. Comparison of fenitrothion and malathion for treatment of plywood and galvanized steel surfaces for control of the red flour beetle and the rusty grain beetle. J. Econ. Entomol. 76:856-863.